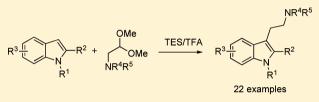
Synthesis of Tryptamine Derivatives via a Direct, One-Pot Reductive Alkylation of Indoles

Marika Righi, Francesca Topi, Silvia Bartolucci, Annalida Bedini, Giovanni Piersanti,* and Gilberto Spadoni

Department of Biomolecular Sciences, University of Urbino, Piazza Rinascimento 6, 61029 Urbino (PU), Italy

Supporting Information

ABSTRACT: An efficient, one-pot reductive alkylation of indoles with *N*-protected aminoethyl acetals in the presence of TES/TFA is reported. It represents the first general method for the direct synthesis of tryptamine derivatives from indoles and nitrogenfunctionalized acetals. This convergent and versatile approach employs safe and inexpensive reagents, proceeds under mild conditions, and tolerates several functional groups. The new



procedure was efficiently applied to a gram-scale synthesis of both luzindole, a reference MT2-selective melatonin receptor antagonist, and melatonin.

T ryptamines are an important class of biologically active compounds whose structural motif is embedded in numerous natural products, including commercial drugs¹ (Figure 1). In addition, tryptamine is a biosynthetic precursor of many alkaloid natural products, including approximately 3000 monoterpene indole alkaloids,² and is frequently used as a chemical building block in the total synthesis of biologically active and pharmaceutically important compounds.³ Thus, there is currently increased interest in tryptamines, and methods for their synthesis are being studied.^{4–8}

A variety of well-documented traditional and modern methods for the synthesis of tryptamines have been described, which can be divided into two broad categories. The first approach involves a multistep sequence for the introduction of the ethylene amine side chain in position 3 of the indole ring. Two of the most important methods in this category utilize an electrophilic aromatic substitution reaction with oxalyl chloride followed by amidation and reduction⁴ or nitroethylation of indoles followed by reduction.⁵ The second approach constructs the pyrrole ring using an annulation method, such as the Fischer⁶ indolization of arylhydrazines with a 4aminobutanal equivalent or the Larock heteroannulation between *o*-haloanilines and an internal alkyne.⁷ Recently, Nicolaou et al. reported a new three-step synthesis of tryptamines starting from N-Boc-protected anilines.⁸ However, some drawbacks of the above methodologies, including the necessary isolation or preparation of reactive intermediates and the use of multistep reaction sequences (one of which requires the use of a strong reducing agent), leave room for improvements.

As part of our ongoing efforts to synthesize melatonin receptor agonists and antagonists,⁹ we recently reported an efficient and selective one-pot reductive amination of anilines for the rapid assembly of functionalized unsymmetrically substituted ethylenediamine derivatives.¹⁰ A convenient alkylat-

ing system for our purpose was N-protected aminoacetaldehyde dimethyl acetal and Et₃SiH/TFA as a reagent combination. We speculated that these reagents and reaction conditions could also be used for the C-3 reductive alkylation of indoles, in order to obtain tryptamine derivatives of pharmaceutical interest in a single step. Although some examples of C-3 reductive alkylation of indoles are known,¹¹ there are no reports describing the use of this protocol for the direct synthesis of tryptamine derivatives. Herein, we describe the optimized conditions, scope and applications for the selective C-3 reductive alkylation of indoles with some protected aminoacetaldehydes (glycinaldehydes). We envisaged that achieving C-3 reductive alkylation of indoles with suitable two-carbon nitrogen-containing electrophiles such as N-protected aminoacetaldehyde dimethyl acetals, would open up promising perspectives to the rapid and practical synthesis of tryptamine and its derivatives.

Our investigation began by screening reaction parameters using 2-methylindole 1a, which is not prone to polymerization, as the model substrate and N-acetylaminoacetaldehyde dimethyl acetal 2 with triethylsilane as the reductant, in DCM (Table 1). As expected, no reaction occurred in the absence of any acid additive (Table 1, entry 1). In agreement with previous reports,¹¹ in the presence of 1.5 equiv of trifluoroacetic acid (TFA), bisindolyl derivative 4 was the major product instead of the desired tryptamine **3a** (Table 1, entry 2). The amount of TFA required in our C-3 reductive alkylation protocol could be critical, as it is necessary to activate different stages of the process. Therefore, we decided to increase the amount of TFA used, and we repeated the reaction using 3 equiv of TFA. As outlined in Table 1 (entry 3), we observed formation of the desired 2-methyl-N-acetamidotryptamine in addition to the bisindolyl derivative 4. Encouraged by this

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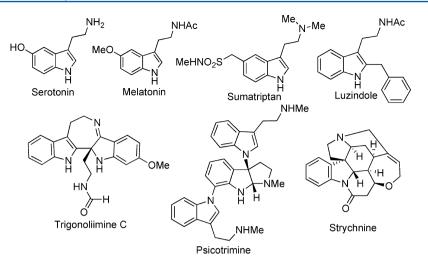


Figure 1. Natural products and pharmaceuticals containing tryptamines.

Table 1. Optimization of the Reaction Parameters

	NHCOCH ₃ NHCOCH ₃							
	1a	2	3 a	4				
						yield ^b (%)		
entry ^a	TFA (equiv)	solvent (0.25 M)	silane	time (h)		4		
1		DCM	TES	48	NR ^c	NR		
2	1.5	DCM	TES	16		91		
3	3	DCM	TES	16	39	45		
4	5	DCM	TES	3	90			
5	5	CHCl ₃	TES	3	85			
6	5	toluene	TES	3	71			
7	5	THF	TES	16	NR	NR		
8	5	Et ₂ O	TES	16	NR	NR		
9	5	dioxane	TES	16	NR	NR		
10	5	CH ₃ CN	TES	16	47			
11	5	DMF	TES	16	36			
12	5	TFE	TES	16	35			
13	5	H ₂ O	TES	16		52		
14	5	DCM	PMHS	16		11		
15	5	DCM	(EtO) ₃ SiH	16		32		
16	5	DCM	TMDSO	16	d			
17	5	DCM	Ph_2SiH_2	16	d			

^aAll reactions were performed at room temperature with 1.1 equiv of acetal, 1.0 equiv of 1a, and 3 equiv of silane in solvent/TFA with no precautions taken to exclude moisture. ^bIsolated yields. ^cNR: no reaction. ^dA complicated crude mixture was observed.

result, we further increased the amount of TFA (5 equiv) to obtain the desired tryptamine derivative 3a as the only reaction product in excellent isolated yield (90%) (Table 1, entry 4). A similar phenomenon was observed in chloroform and toluene, although the selectivity and yields were lower than in DCM. All our attempts to improve the yields or the selectivity outcome by changing the solvent were unsuccessful (Table 1, entries 7-13). Decreased yields of the desired tryptamine derivative were observed in trifluoroethanol, DMF, and acetonitrile, and no reaction occurred with ethereal solvents such as diethyl ether, THF, and dioxane. Only the bisindolyl adduct 4 was observed in water. It appears that decreasing the nucleophilicity of the medium increases the acidity of the system, thus promoting the

reaction. In addition to Et₃SiH, other silanes such as Ph₂SiH₂, (EtO)₃SiH, polymethylhydrosiloxane (PMHS), and tetramethyldisiloxane (TMDS) were also tested as reductants, but none of them gave appreciable results (Table 1, entries 14–17).

Once reliable conditions to synthesize 2-methyl-N-acetamidotryptamine were established, we next investigated the scope of this reaction using indoles with different substitution patterns as summarized in Table 2.

A variety of 2-alkyl/aryl-substituted indoles were alkylated smoothly in good yields, regardless of the length or steric hindrance of the C-2 substituent (Table 2, entries 1-6). Interestingly, it was also possible to perform C-3 reductive alkylation of N ω -acetyl isotryptamine $1g^{12}$ to give the

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Note

Table 2. Scope of Indoles 1a-t in the C-3 Reductive Alkylation with 2

					NHCOCH₃		
			ŅHCOCH ₃				
$R^3 \frac{1}{11}$	R^2	+ MeO	TES/	FFA → R ³ …		₹ ²	
	\sim_{N}		CH ₂ C OMe	l₂ rt	N		
1a	R ¹		2		R ¹		
14-1		2		3a-g, 3j-t			
, а	1.	\mathbb{R}^1	R^2	R ³	time	yield ^b	
entry ^a	product				(h)	(%)	
1	3a	Н	Me	Н	3	90	
2	3b	Н	But	Н	3	70	
3	3c	Н	cyclopentyl	Н	6	76	
4	3d	Н	<i>t</i> -But	Н	3	66	
5	3e	Н	Bn	Н	4	74	
6	3f	Н	Ph	Н	4	97	
7	3g	Н	CH ₂ CH ₂ NHAc	Н	3	60	
8	3h	Н	СООН	Н		NR^d	
9	3i	Н	COOMe	Н		NR	
10	3j	Н	Me	5-Me	3	85	
11	3k	Н	Me	5-Cl	3	87	
12	31	Н	Me	5-Bpin	3	70	
13	3m	Н	Me	5-OMe	2	63 ^c	
14	3n	Н	Ph	6-OMe	2	43 ^c	
15	30	Me	Me	Н	3	77	
16	3p	Me	Н	Н	4	50	
17	3q	Bn	Н	Н	16	65	
18	3r	Bn	Н	5-OMe	2	56 ^c	
19	3s	PMB	Н	Н	16	45	
20	3t	Allyl	Н	Н	16	40	

^{*a*}Reaction conditions: 2 (1 mmol), Ia-t (1.1 mmol), TFA (5 mmol), and TES (3 mmol) in DCM (0.25 M) at rt for 3–60 h. ^{*b*}Yield after flash chromatography. ^{*c*}DCE at 60 °C. ^{*d*}NR: no reaction.

interesting (bis-ethylamino)indole derivative 3g as a single product in good yield (60%) (Table 2, entry 7); the same compound 3g could be theoretically obtained by reacting N ω acetyltryptamine under the same reaction conditions, but these attempts gave only polymerization side products. On the contrary, no reactivity was observed with indoles bearing an electron-withdrawing group in the C-2 position, such as 2indolecarboxylic acid or its ester (Table 2, entries 8 and 9), even if higher reaction temperatures and prolonged reaction times were used. A possible reason is that the presence of an electron-withdrawing group might render the indoles highly electron-deficient, preventing the electrophilic alkylation. A similar lack of reactivity was observed with N1-acetyl- and N1sulfonylindoles (data not shown). Our synthetic protocol worked well for indoles with both moderately electrondonating (i.e., methyl, entry 10) and electron-withdrawing (i.e., chloro, entry 11) 5-substituents giving the corresponding tryptamines in high yields. However, when 5-methoxy-2methylindole reacted with acetal 2, only 19% yield was obtained. However, increasing the reaction temperature to 60 °C led to a consistent improvement in the yield of the desired tryptamine (Table 2, entry 13). Remarkably, the proposed C-3 reductive alkylation also tolerates the presence of a boronate ester in the indole ring, offering unique, site-specific handles that can be utilized in cross-coupling methodology for further functionalization of tryptamines (Table 2, entry 12). Encouraged by these results, we applied the reaction conditions to 2-unsubstituted indole substrate. Gratifingly, the above

indole C-3 reductive alkylation strategy could also be extended to *N*-alkylated indoles 10-t,¹³ independent of the presence of a C-2 indole substituent to give the corresponding desired tryptamines 30-t, although in slightly lower yields (45–77% yield) and requiring longer reaction times (Table 2, entries 16–20).

Unfortunately, the indole does not afford the desired $N\omega$ acetyltryptamine. The shortcomings associated with completely unsubstituted indoles in the reductive alkylation reaction are likely a manifestation of increased reactivity of the product toward further reaction with electrophiles present in the reaction mixture, giving a complicated mixture of dimer and oligomer products.¹⁴

To further expand the potential of this new one-pot singlestep synthesis of tryptamine derivatives, we also investigated an orthogonal set of easily removable N-protected aminoalkyl acetals.¹⁵ The reaction proceeded well in all cases (Table 3,

Table 3. Scope of Acetals 5–9 in the Reductive Alkylation of 1a

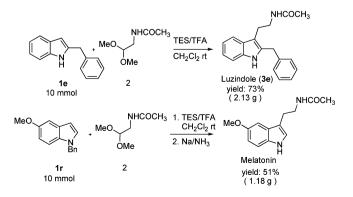
	Me + MeO NHF Me + MeO OMe a 5-9	$\frac{\text{TES/TFA}}{\text{CH}_2\text{CI}_2 \text{ rt}}$	- N H 3u-y	NHPg / -Me
entry ^a	protecting group (Pg)	time (h)	yield ^{b} (%)	product
1	Cbz (5)	16	60	3u
2	Ftal (6)	16	78	3v
3	Ts (7)	16	50	3w
4	$\text{COCF}_3(8)$	16	89	3x
5	Н (9)	16	NR ^c	$3y^d$

^{*a*}Reaction conditions: 5-9 (1 mmol), 1a (1.1 mmol), TFA (5 mmol), and TES (3 mmol) in DCM (0.25 M) at rt for 3-16 h. ^{*b*}Yield after flash chromatography. ^{*c*}NR: no reaction. ^{*d*}From 3x using base.

entries 1-4), although more slowly. Interestingly, the *N*-trifluoroacetyl protecting group proved to be well tolerated to the reaction conditions and easily removed by simple basic hydrolysis, directly on the crude reaction mixture, to give tryptamine **3y** in very high yield;¹⁶ the same compound was not obtained by using the simple amino acetal **9**.

A wide range of synthetic applications of this procedure can be envisaged. For instance, the method was extended to the practical gram-scale synthesis of luzindole (Scheme 1), a widely used and expensive MT_2 melatonin receptor antagonist.

Scheme 1. Multigram-Scale Reductive Alkylation for the Synthesis of Luzindole and Melatonin



Although it constitutes a standard reference compound for pharmacologists in the melatonin field, the reported multistep preparation sequences are laborious and give poor overall vields.¹⁷ The single step gram-scale preparation reaction is conveniently carried out at room temperature under an air atmosphere with the inexpensive and easily handled starting material 2-benzylindole in good yield. In order to show the viability of this method, we also prepared 2-unsubstituted tryptamine derivatives in gram scale. The treatment of readily available N-benzyl-5-methoxyindole with acetal 2, followed by reductive removal of the N1-benzyl group by Na/NH₃, afforded, in acceptable yields, N-acetyl-5-methoxytryptamine (melatonin), a neurohormone mainly secreted by the pineal gland and known to play a key role in regulating the body's circadian rhythms, including the sleep-wake cycle and mood.¹⁸ This chemistry can in principle be easily adapted to provide access to various analogues of 5-alkoxy- and/or 5-hydroxytryptamine, which may not be readily available otherwise.

In summary, a new, direct, one-pot method for the synthesis of tryptamine derivatives has been developed, via C-3 reductive alkylation of indoles and readly available protected glycinaldehyde acetals. Protected glycinaldehyde acetals have shown to be useful two-carbon nitrogen containing electrophiles and could be utilized to obtain products incorporating the amino ethane fragment. The present approach constitutes one of the most efficient, practical, and straightforward methods for a variety of substituted tryptamines. This powerfully simplified reaction has enabled the gram-scale synthesis of two important and biologically active tryptamine derivatives: the neurohormone melatonin and the reference melatonin receptor antagonist luzindole. Given the simplicity, efficiency, and functional group tolerance, this general method is expected to find wide applications in chemical synthesis in general and in the construction of tryptamine-based complex molecules.

EXPERIMENTAL SECTION

General Information. All reactions were run in round-bottom flasks. Column chromatography purifications were performed in flash conditions using 230–400 mesh silica gel. Analytical thin-layer chromatography (TLC) was carried out on silica gel plates (silica gel 60 F_{254}) that were visualized by exposure to ultraviolet light and an aqueous solution of cerium ammonium molybdate (CAM). ¹H NMR and ¹³C NMR spectra were recorded at 200/50 MHz on spectrometer using CDCl₃ or DMSO- d_6 as solvent. Chemical shifts (scale) are reported in parts per million (ppm) relative to the central peak of the solvent. Coupling constants (*J* values) are given in hertz (Hz). ESI-MS spectra were taken on an LC–MS instrument. Only molecular ions (M + 1) are given for the ESI-MS analysis. Absorbances are reported in cm^{-1} for the IR analysis. Melting points were determined on a capillary melting point apparatus and are uncorrected. Elemental analyses were within ±0.4 of the theoretical values (C, H, N).

General Procedure for the Reductive Alkylation of Indoles. A solution of suitable indole (1 mmol) and acetal (1.1 mmol) in DCM (2 mL) was added to a solution of trifluoroacetic acid (TFA, 5 mmol) and triethylsilane (TES, 3 mmol) in DCM (2 mL), and the resulting mixture was stirred at room temperature for 3-16 h. The reaction was cooled at 0 °C and carefully neutralized with saturated solution of NaHCO₃ and diluted with DCM. The two phases were separated, and the aqueous layer was extracted three times with DCM (20 mL). The combined organic phase was washed with brine, dried over Na₂SO₄, filtered, and evaporated on rotary evaporator. The residue was purified by flash chromatography on silica gel affording desired compounds.

N-[2-(2-Methyl-1H-indol-3-yl)ethyl]acetamide (3a). Purification by silica gel column chromatography (EtOAc) gave the product in 90% yield (194 mg): yellow solid; mp 83–85 °C (EtOAc); MS (ESI) 217 $[M + H]^+$; IR (film, cm⁻¹) 1620; ¹H NMR (CDCl₃) δ 1.89 (s, 3H), 2.36 (s, 3H), 2.91 (t, *J* = 6.5 Hz, 2H), 3.49 (dt, $J_1 \approx J_2 = 6.5$ Hz, 2H), 5.75 (bs, 1H), 7.05–7.17 (m, 2H), 7.25–7.29 (m, 1H), 7.47–7.51 (m, 1H), 8.44 (bs, 1H). ¹³C NMR (CDCl₃) δ 11.5, 23.3, 24.1, 40.1, 108.3, 110.4, 117.7, 119.3, 121.1, 128.6, 132.1, 135.3, 170.3. Anal. Calcd for C₁₃H₁₆N₂O (216): C, 72.19; H, 7.46; N, 12.95. Found: C, 72.10; H, 7.18; N, 12.94. Data was consistent with that reported in the literature.^{18a}

N-[2-(2-Butyl-1H-indol-3-yl)ethyl]acetamide (**3b**). Purification by silica gel column chromatography (cyclohexane/EtOAc 1:1) gave the product in 70% yield (181 mg): yellowish oil; ESI MS (*m*/*z*) 259 [M + H]⁺; IR (cm⁻¹, film) 1620; ¹H NMR (CDCl₃) δ 0.96 (t, *J* = 7 Hz, 3H), 1.35–1.50 (m, 2H), 1.58–1.73 (m, 2H), 1.91 (s, 3H), 2.75 (t, *J* = 7 Hz, 2H), 2.93 (t, *J* = 6.5 Hz, 2H), 3.54 (dt, *J*₁ ≈ *J*₂ = 6.5 Hz, 2H), 5.53 (bs, 1H), 7.07–7.19 (m, 2H), 7.27–7.34 (m, 1H), 7.51–7.55 (m, 1H), 7.98 (bs, 1H); ¹³C NMR (CDCl₃) δ 13.9, 22.6, 23.3, 24.1, 25.8, 32.2, 40.2, 107.9, 110.6, 117.9, 119.2, 121.1, 128.5, 135.4, 136.9, 170.3. Anal. Calcd for C₁₆H₂₂N₂O (258): C, 74.38; H, 8.58; N, 10.84. Found: C, 74.29; H, 8.45; N, 10.69. Data was consistent with that reported in the literature.^{18a}

N-(2-(2-Cyclopentyl-1*H*-indol-3-yl)ethyl)acetamide (**3c**). Purification by silica gel column chromatography (cyclohexane/EtOAc 1:1) gave the product in 76% yield (205 mg): off-white solid; mp 147–149 °C (Et₂O); ESI MS (*m*/*z*) 271 [M + H]⁺; IR (cm⁻¹, film) 1654; ¹H NMR (CDCl₃) δ 1.60–2.09 (m, 11H), 2.96 (t, *J* = 6.5 Hz, 2H), 3.22–3.39 (m, 1H), 3.54 (dt, *J*₁ ≈ *J*₂ = 6.5 Hz, 2H), 5.62 (bs, 1H), 7.06–7.19 (m, 2H), 7.27–7.35 (m, 1H), 7.50–7.54 (m, 1H), 8.19 (bs, 1H); ¹³C NMR (CDCl₃) δ 23.4, 24.2, 25.8, 33.6, 36.7, 40.2, 107.8, 110.6, 117.9, 119.3, 121.2, 128.5, 135.4, 139.8, 170.0. Anal. Calcd for C₁₇H₂₂N₂O (270): C, 75.52; H, 8.20; N, 10.36. Found: C, 75.56; H, 8.39; N, 10.64.

N-(2-(2-tert-Butyl-1*H*-indol-3-yl)ethyl)acetamide (**3***d*). Purification by silica gel column chromatography (cyclohexane/EtOAc 1:1) gave the product in 66% yield (170 mg): amorphous off-white solid; ESI MS (*m*/*z*) 259 [M + H]⁺; IR (cm⁻¹, film) 1647; ¹H NMR (CDCl₃) δ 1.50 (s, 9H), 1.95 (s, 3H), 3.14 (t, *J* = 7 Hz, 2H), 3.59 (dt, *J*₁ ≈ *J*₂ = 7 Hz, 2H), 5.84 (bs, 1H), 7.06–7.20 (m, 2H), 7.32–7.36 (m, 1H), 7.55–7.60 (m, 1H), 8.26 (bs, 1H); ¹³C NMR (CDCl₃) 23.4, 25.4, 30.7, 33.0, 40.5, 106.7, 110.5, 118.0, 119.3, 121.2, 129.7, 134.1, 143.0, 170.2. Anal. Calcd for C₁₆H₂₂N₂O (258): C, 74.38; H, 8.58; N, 10.84. Found: C, 74.45; H, 8.63; N, 11.12.

N-[2-(2-Benzyl-1H-indol-3-yl)ethyl]acetamide (**3e**). The reaction was run on 10 mmol (2.07 g) scale. Purification by silica gel column chromatography (cyclohexane/EtOAc 1:1) gave the product in 74% yield (2.16 g): white solid; mp 103−104 °C (EtOAc); ESI MS (*m*/*z*) 293 [M + H]⁺; IR (cm⁻¹, film) 1630; ¹H NMR (CDCl₃) δ 1.80 (*s*, 3H), 3.01 (t, *J* = 6.5 Hz, 2H), 3.54 (dt, *J*₁ ≈ *J*₂ = 6.5 Hz, 2H), 4.13 (*s*, 2H), 5.48 (bs, 1H), 7.08−7.38 (m, 8 H), 7.55−7.59 (m, 1H), 7.84 (bs, 1H); ¹³C NMR (CDCl₃) δ 23.2, 24.2, 32.2, 40.1, 109.4, 110.7, 118.2, 119.5, 121.6, 126.8. 128.4, 128.5, 128.9, 134.3, 135.7, 138.7, 170.1. Anal. Calcd for C₁₉H₂₀N₂O (292): C, 78.05; H, 6.89; N, 9.58. Found: C, 78.14; H, 6.75; N, 9.64. Data was consistent with that reported in the literature.^{17b}

N-[2-(2-Phenyl-1H-indol-3-yl)ethyl]acetamide (**3f**). Purification by silica gel column chromatography (cyclohexane/EtOAc 1:1) gave the product in 97% yield (270 mg): yellow solid; mp 114−116 °C (hexane); ESI MS (*m*/*z*) 279 [M + H]⁺; IR (cm⁻¹, film) 1620; ¹H NMR (CDCl₃) δ 1.77 (s, 3H), 3.14 (t, *J* = 7 Hz, 2H), 3.56 (dt, *J*₁ ≈ *J*₂ = 7 Hz, 2H), 5.52 (bs, 1H), 7.13−7.27 (m, 2 H), 7.35−7.68 (m, 7H), 8.37 (bs, 1H); ¹³C NMR (CDCl₃) δ 23.1, 24.4, 40.2, 109.7, 111.1, 118.9, 119.8, 122.4, 127.9, 128.0, 129.0, 133.0, 135.4, 136.0, 170.2. Anal. Calcd for C₁₈H₁₈N₂O (278): C, 77.67; H, 6.52; N, 10.06. Found: C, 77.65; H, 6.45; N, 10.21. Data was consistent with that reported in the literature.^{18a}

N,*N'*-[2,2'-(1*H*-Indole-2,3-diyl)bis(ethane-2,1-diyl)]diacetamide (**3g**). Purification by silica gel column chromatography (EtOAc/MeOH 9:1) gave the product in 60% yield (172 mg): amorphous off-white solid; ESI MS (*m*/*z*) 288 [M + H]⁺; IR (cm⁻¹, film) 1640; ¹H NMR (DMSO- d_6) δ 1.79 (s, 3H), 1.81 (s, 3H), 2.73–2.86 (m, 4H), 3.17–3.40 (m, 4H), 6.90–7.04 (m, 2H), 7.24–7.28 (m, 1H), 7.45–7.48 (m, 1H), 7.90–7.95 (m, 2H), 10.76 (s, 1H); ¹³C NMR (DMSO- d_6) δ 23.0, 23.1, 24.7, 26.5, 108.9, 111.1, 118.1, 118.6, 120.7, 128.6,

134.3, 136.0, 169.5, 169.7. Anal. Calcd for $C_{16}H_{21}N_3O_2$ (287): C, 66.88; H, 7.37; N, 14.62. Found: C, 66.91; H, 7.56; N,14.91.

N-[2-(2,5-Dimethyl-1H-indol-3-yl)ethyl]acetamide (**3***j*). Purification by silica gel column chromatography (EtOAc) gave the product in 85% yield (196 mg): red viscous oil; ESI MS (*m*/*z*) 231 [M + H]⁺; IR (cm⁻¹, film) 1650, 2920, 3280; ¹H NMR (CDCl₃) δ 1.91 (s, 3H), 2.37 (s, 3H), 2.46 (s, 3H), 2.89 (t, *J* = 6.5 Hz, 2H), 3.50 (dt, *J*₁ ≈ *J*₂ = 6.5 Hz, 2H), 5.58 (bs, 1H), 6.97 (d, *J* = 8 Hz, 1H), 7.18 (d, *J* = 8 Hz, 1H), 7.28–7.29 (m, 1H), 7.97 (bs, 1H); ¹³C NMR (CDCl₃) δ 11.6, 21.5, 23.3, 24.1, 40.1, 107.9, 110.1, 117.6, 122.6, 128.5, 128.8, 132.2, 133.6, 170.2. Anal. Calcd for C₁₄H₁₈N₂O (230): C, 73.01; H, 7.88; N, 12.16. Found: C, 73.03; H, 7.81; N, 12.22 .

N-(2-(5-*Chloro-2-methyl-1H-indol-3-yl)ethyl)acetamide* (**3***k*). Purification by silica gel column chromatography (EtOAc) gave the product in 87% yield (218 mg): pink solid; mp 121–125 °C (Et₂O); ESI MS (*m*/*z*) 251 [M + H]⁺; IR (cm⁻¹, film) 1630; ¹H NMR (CDCl₃) δ 1.92 (s, 3H), 2.34 (s, 3H), 2.85 (t, *J* = 7 Hz, 2H), 3.44 (dt, *J*₁ ≈ *J*₂ = 7 Hz, 2H), 5.82 (bs, 1H), 7.01–7.18 (m, 1H), 7.43 (1H), 8.66 (bs, 1H); ¹³C NMR (CDCl₃) δ 23.1, 24.4, 40.2, 109.7, 111.1, 118.9, 119.8, 122.4, 127.9, 128.0, 129.0, 133.0, 135.4, 136.0, 170.2. Anal. Calcd for C₁₃H₁₅ClN₂O (250): C, 62.28; H, 6.03; N, 11.17. Found: C, 62.35; H, 6.28; N, 11.51.

N-(2-(2-*Methyl*-5-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1*H*-indol-3-yl)ethyl)acetamide (**3***l*). Purification by silica gel column chromatography (EtOAc) gave the product in 70% yield (239 mg): amorphous off-white solid; ESI MS (*m*/*z*) 343 [M + H]⁺; IR (cm⁻¹, film) 1658; ¹H NMR (CDCl₃) δ 1.37 (s, 12H), 1.91 (s, 3H), 2.37 (s, 3H), 2.91 (t, *J* = 6.5 Hz, 2H), 3.47 (dt, *J*₁ ≈ *J*₂ = 6.5 Hz, 2H), 5.55 (bs, 1H), 7.25–7.29 (m, 1H), 7.59 (d, *J* = 8 Hz, 1H), 8.00 (s, 1H), 8.21 (bs, 1H); ¹³C NMR (CDCl₃) δ 11.5, 23.3, 24.0, 24.9, 40.3, 83.5, 109.0, 109.9, 125.4, 127.5, 128.3, 132.2, 137.5, 170.2 (carbon adjacent to boron was not observed). Anal. Calcd for C₁₉H₂₇BN₂O₃ (342): C, 66.68; H, 7.95; N, 8.19. Found: C, 66.65; H, 7.98; N, 8.31.

N-[2-(5-*Methoxy*-2-*methyl*-1*H*-*indol*-3-*yl*)*ethyl*]*acetamide* (**3***m*). Purification by silica gel column chromatography (cyclohexane/ EtOAc 3:7) gave the product in 63% yield (155 mg): brownish oil; ESI MS (*m*/*z*) 247 [M + H]⁺; IR (cm⁻¹, film)1630; ¹H NMR (CDCl₃) δ 1.91 (*s*, 3H), 2.36 (*s*, 3H), 2.88 (*t*, 2H, *J* = 6.5 Hz), 3.49 (d*t*, 2H, *J*₁ ≈ *J*₂ = 6.5 Hz), 3.85 (*s*, 3H), 5.58 (bs, 1H), 6.76–6.81 (m, 1H), 6.95–6.97 (m, 1H), 7.15–7.19 (m, 1 H), 7.94 (bs, 1H); ¹³C NMR (CDCl₃) δ 11.6, 23.3, 24.1, 40.0, 56.0, 100.1, 108.2, 110.7, 111.1, 129.0, 130.4, 133.0, 154.0, 170.2. Anal. Calcd for C₁₄H₁₈N₂O₂ (246): C, 68.27; H, 7.37; N, 11.37. Found: C, 68.25; H, 7.45; N, 11.22. Data was consistent with that reported in the literature.^{6d}

N-[2-(6-*Methoxy*-2-*phenyl*-1*H*-*indol*-3-*yl*)*ethyl*]*acetamide* (**3***n*). Purification by silica gel column chromatography (cyclohexane/ EtOAc 4:6) gave the product in 43% yield (78 mg): brownish oil; ESI MS (*m*/*z*) 309 [M + H]⁺; IR (cm⁻¹, film) 1625; ¹H NMR (CDCl₃) δ 1.76 (s, 3H), 3.09 (t, 2H, *J* = 7.0 Hz), 3.53 (dt, 2H, *J*₁ ≈ *J*₂ = 7.0 Hz), 3.86 (s, 3H), 5.58 (bs, 1H), 6.79–6.90 (m, 2H), 7.28–7.56 (m, 6H), 8.43 (bs, 1H); ¹³C NMR (CDCl₃) δ 23.2, 24.5, 40.2, 55.7, 94.6, 109.7, 119.6, 123.5, 127.5, 127.7, 129.0,133.1, 134.1,136.8, 156.8, 170.2. Anal. Calcd for C₁₉H₂₀N₂O₂ (308): C, 74.00; H, 6.54; N, 9.08. Found: C, 74.11; H, 6.39; N, 9.36.

N-[2-(1,2-Dimethyl-1*H*-indol-3-yl)ethyl]acetamide (**30**). Purification by silica gel column chromatography (cyclohexane/EtOAc 3:7) gave the product in 77% yield (177 mg): pink solid; mp 101−104 °C (Et₂O); ESI MS (*m*/*z*) 231 [M + H]⁺; IR (cm⁻¹, film) 1650; ¹H NMR (CDCl₃) δ 1.91 (s, 3H), 2.39 (s, 3H), 2.96 (t, 2H, *J* = 6.5 Hz), 3.49 (dt, 2H, *J*₁ ≈ *J*₂ = 6.5 Hz), 3.68 (s, 3H), 5.65 (bs, 1H), 7.07−7.31 (m, 3H), 7.51−7.55 (m, 1H); ¹³C NMR (CDCl₃) δ 10.2, 23.4, 24.4, 29.6, 40.3, 107.7, 108.7, 117.7, 119.0, 120.8, 127.7, 133.8, 136.6, 170.0. Anal. Calcd for C₁₄H₁₈N₂O (230): C, 73.01; H, 7.88; N, 12.16. Found: C, 73.11; H, 7.92; N, 12.01. Data was consistent with that reported in the literature.^{6d}

N-[2-(1-Methyl-1H-indol-3-yl)ethyl]acetamide (3p). Purification by silica gel column chromatography (EtOAc) gave the product in 50% yield (108 mg): yellowish oil; ESI MS (m/z) 217 [M + H]⁺; IR (cm⁻¹, film) 1630; ¹H NMR (CDCl₃) δ 1.94 (s, 3H), 2.97 (t, J = 7 Hz, 2 H), 3.59 (dt, $J_1 \approx J_2 = 7$ Hz, 2H), 3.78 (s, 3H), 5.64 (bs, 1H), 6.90

(s, 1H), 7.10–7.35 (m, 3H), 7.59–7.63 (m, 1H); ¹³C NMR (CDCl₃) δ 11.6, 23.3, 24.1, 40.1, 108.4, 110.4, 117.7, 119.3, 121.1, 128.6, 132.1, 135.3, 170.2. Anal. Calcd for C₁₃H₁₆N₂O (216): C, 72.19; H, 7.46; N, 12.95. Found: C, 72.08; H, 7.55; N, 12.81. Data was consistent with that reported in the literature.¹³

N-[2-(1-Benzyl-1H-indol-3-yl)ethyl]acetamide (**3q**). Purification by silica gel column chromatography (EtOAc) gave the product in 65% yield (190 mg): yellowish oil; ESI MS (*m*/*z*) 293 [M + H]⁺; IR (cm⁻¹, film) 1630; ¹H NMR (CDCl₃) δ 1.92 (s, 3H), 2.99 (t, *J* = 7 Hz, 2H), 3.59 (dt, *J*₁ ≈ *J*₂ = 7 Hz, 2H), 5.30 (s, 2H), 5.62 (bs, 1H), 6.98 (s, 1H), 7.11–7.33 (m, 8H), 7.62–7.66 (m, 1H); ¹³C NMR (CDCl₃) δ 23.3, 25.3, 40.0, 49.9, 109.8, 112.3, 119.0, 119.3, 122.0, 126.1, 126.9, 127.7, 128.1, 128.8, 136.8, 137.6, 170.1. Anal. Calcd for C₁₉H₂₀N₂O (292): C, 78.05; H, 6.89; N, 9.58. Found: C, 78.14; H, 6.95; N, 9.39. Data was consistent with that reported in the literature.¹³

N-[2-(1-Benzyl-5-methoxy-1*H*-indol-3-yl)ethyl]acetamide (**3***r*). The reaction was run on 10 mmol (2.37 g) scale. Purification by silica gel column chromatography (EtOAc) gave the product in 56% yield (1.81 g): off-white solid; mp 115 °C (EtOAc/hexane); ESI MS (*m*/*z*) 323 [M + H]⁺; IR (cm⁻¹, film) 1641; ¹H NMR (CDCl₃) δ 1.93 (s, 3H), 2.92 (t, *J* = 7 Hz, 2H), 3.59 (dt, *J*₁ ≈ *J*₂ = 7 Hz, 2H), 3.87 (s, 3H), 5.24 (s, 2H), 5.73 (bs, 1H), 6.85–6.95 (m, 2H), 7.08–7.20 (m, 4H), 7.29–7.32 (m, 3H); ¹³C NMR (CDCl₃) δ 23.4, 25.3, 40.0, 49.4, 55.3, 109.8, 112.1, 114.1, 119.0, 119.2, 121.9, 126.0, 128.1, 128.3, 129.5, 136.7, 159.1, 170.1. Anal. Calcd for C₂₀H₂₂N₂O₂ (322): C, 74.51; H, 6.88; N, 8.69. Found: C, 74.41; H, 6.99; N, 8.59.

N-[2-(1-(4-*Methoxybenzyl*)-1*H*-*indol*-3-*yl*)*ethyl*]*acetamide* (**3s**). Purification by silica gel column chromatography (EtOAc) gave the product in 45% yield (145 mg): amorphous off-white solid; ESI MS (*m*/*z*) 323 [M + H]⁺; IR (cm⁻¹, film) 1626; ¹H NMR (CDCl₃) δ 1.92 (s, 3H), 2.98 (t, *J* = 6 Hz, 2H), 3.59 (dt, *J*₁ ≈ *J*₂ = 6 Hz, 2H), 3.79 (s, 3H), 5.22 (s, 2H), 5.58 (bs, 1H), 6.82–6.87 (m, 2H), 6.95 (s, 1H), 7.07–7.35 (m, 5H), 7.60–7.64 (m, 1H); ¹³C NMR (CDCl₃) δ 23.4, 25.3, 40.0, 49.4, 55.3, 109.8, 112.1, 114.1, 119.0, 119.2, 121.9, 126.0, 128.1, 128.3, 129.5, 136.7, 159.1, 170.1. Anal. Calcd for C₂₀H₂₂N₂O₂ (322): C, 74.51; H, 6.88; N, 8.69. Found: C, 74.43; H, 7.11; N, 8.81.

N-[2-(1-Allyl-1H-indol-3-yl)ethyl]acetamide (**3t**). Purification by silica gel column chromatography (EtOAc) gave the product in 40% yield (97 mg): amorphous off-white solid; ESI MS (*m*/*z*) 243 [M + H]⁺; IR (cm⁻¹, film) 1650; ¹H NMR (CDCl₃) δ 1.94 (s, 3H), 2.98 (t, *J* = 6 Hz, 2H), 3.60 (dt, *J*₁ ≈ *J*₂ = 6 Hz, 2H), 4.69–4.72 (m, 2H), 5.06–5.25 (m, 2H), 5.77 (bs, 1H), 5.90–6.09 (m, 1H), 6.96 (s, 1H), 7.10–7.35 (m, 3H), 7.60–7.64 (m, 1H); ¹³C NMR (CDCl₃) δ 23.4, 25.2, 40.0, 48.7, 109.7, 112.0, 117.3, 118.9, 119.1, 121.8, 125.7, 128.0, 133.5, 136.6, 170.1. Anal. Calcd for C₁₅H₁₈N₂O (242): C, 74.35; H, 7.49; N, 11.56. Found: C, 74.26; H, 7.24; N, 11.42.

Benzyl 2-(2-Methyl-1H-indol-3-yl)ethylcarbamate (**3u**). Purification by silica gel column chromatography (cyclohexane/EtOAc 8:2) gave the product in 60% yield (185 mg): yellowish oil; ESI MS (*m*/*z*) 309 [M + H]⁺; IR (cm⁻¹, film) 1700, 2930, 3320; ¹H NMR (CDCl₃) δ 2.34 (s, 3H), 2.95 (t, *J* = 7 Hz, 2H), 3.48 (dt, *J*₁ \approx *J*₂ = 7 Hz, 2H), 4.91 (bs, 1H), 5.15 (s, 2H), 7.08–7.55 (m, 9H), 8.03 (bs, 1H); ¹³C NMR (CDCl₃) δ 11.5, 24.6, 41.5, 66.6, 108.2, 110.4, 117.8, 119.3, 121.1, 124.2, 128.1, 128.5, 130.5, 132.1, 135.3, 136.7, 156.5. Anal. Calcd for C₁₉H₂₀N₂O₂ (308): C, 74.00; H, 6.54; N, 9.08. Found: C, 74.08; H, 6.52; N, 9.11.

2-[2-(2-Methyl-1H-indol-3-yl)ethyl]isoindolyn-1,3-dione (**3v**). Purification by silica gel column chromatography (cyclohexane/EtOAc 8:2) gave the product in 78% yield (238 mg): pale yellow solid; mp 171–174 °C (hexane); ESI MS (m/z) 305 [M + H]⁺; IR (cm⁻¹, film) 1680; ¹H NMR (CDCl₃) δ 2.43 (s, 3H), 3.04–3.12 (m, 2H), 3.87–3.95 (m, 2H), 7.05–7.15 (m, 2H), 7.24–7.27 (m, 1H), 7.64–7.87 (m, 6H); ¹³C NMR (CDCl₃) δ 11.5, 23.5, 38.3, 107.8, 110.3, 117.9, 119.4, 121.1, 123.1, 128.6, 132.0, 132.2, 133.9, 135.2, 168.4. Anal. Calcd for C₁₉H₁₆N₂O₂ (304): C, 74.98; H, 5.30; N, 9.20. Found: C, 75.03; H, 5.53; N, 8.93.

4-Methyl-N-[2-(2-methyl-1H-indol-3-yl)ethyl]benzenesulfonamide (3w). Purification by silica gel column chromatography (cyclohexane/EtOAc 7:3) gave the product in 50% yield (164 mg): brownish solid; mp 135–137 °C (hexane); ESI MS

(m/z) 329 [M + H]⁺; IR (cm⁻¹, film) 1654; ¹H NMR (CDCl₃) δ 2.34 (s, 3H), 2.41 (s, 3H), 2.90 (t, *J* = 6 Hz, 2H), 3.22 (dt, *J*₁ ≈ *J*₂ = 6 Hz, 2H), 4.46 (t, *J* = 6 Hz, 1H), 6.99–7.33 (m, 6H), 7.61–7.65 (m, 2H), 7.92 (bs, 1H); ¹³C NMR (CDCl₃) δ 11.8, 21.6, 24.7, 43.3, 107.1, 110.5, 117.7, 119.5, 121.4, 127.1, 128.2, 129.7, 132.6, 135.4, 136.9, 143.4. Anal. Calcd for C₁₈H₂₀N₂O₂S (328): C, 65.83; H, 6.14; N, 8.53. Found: C, 65.75; H, 6.09; N, 8.46. Data was consistent with that reported in the literature.^{6d}

2,2,2-Trifluoro-N-[2-(2-methyl-1H-indol-3-yl)ethyl]acetamide (**3x**). Purification by silica gel column chromatography (cyclohexane/ EtOAc 7:3) gave the product in 89% yield (240 mg): brownish solid; mp 98–102 °C (hexane); ESI MS (m/z) 271 [M + H]⁺; IR (cm⁻¹, film) 1630; ¹H NMR (CDCl₃) δ 2.39 (s, 3H), 3.01 (t, *J* = 6.5 Hz, 2H), 3.63 (dt, $J_1 \approx J_2$ = 6.5 Hz, 2H), 6.50 (bs, 1H), 7.10–7.34 (m, 3H), 7.49–7.53 (m, 1H), 8.01 (bs, 1H); ¹³C NMR (CDCl₃) δ 11.3, 23.5, 40.3, 107.1, 110.6, 115.9 (*J* = 285 Hz), 117.5, 119.6, 121.4, 128.2, 132.4, 135.4, 157.3 (*J* = 35 Hz). Anal. Calcd for C₁₃H₁₃F₃N₂O (270): C, 57.78; H, 4.85; N, 10.37. Found: C, 57.65; H, 4.79; N, 10.26. Data was consistent with that reported in the literature.^{6d}

2-(2-Methyl-1H-indol-3-yl)ethanamine (**3y**). A solution of **3x** (1 mmol) and potassium carbonate (5 mmol) in MeOH (20 mL)/H₂O (1.5 mL) was heated to reflux for 2 h. After removal of volatiles under reduced pressure, to the residue was added water and the mixture extracted three times with dichloromethane. The combined organic phases were washed with brine, dried over sodium sulfate, filtered, and concentrated. The residue was purified by silica gel flash chromatography (dichloromethane/methanol 95:5): yield 75% (131 mg); mp 95 °C; ESI MS (m/z) 175 [M + H]⁺, 158 [M + H – NH₃]⁺; ¹H NMR (CDCl₃) δ 1.83 (bs, 2H), 2.38 (s, 3H), 2.83–3.02 (m, 4H), 7.06–7.17 (m, 2H), 7.25–7.29 (m, 1H), 7.50–7.54 (m, 1H), 8.14 (bs, 1H); ¹³C NMR (CDCl₃) δ 11.7, 28.1, 42.5, 108.8, 110.3, 117.9, 119.1, 120.9, 128.7, 132.0, 135.4. Anal. Calcd for C₁₁H₁₄N₂ (174): C, 75.82; H, 8.10; N, 16.08. Found: C, 75.79; H, 8.17; N, 16.18. Data was consistent with that reported in the literature.¹⁶

N-[*2*,2-*bis*(2-*Methyl*-1*H*-3-*yl*)*ethyl*]*acetamide* (*4*). Compound 4 was isolated as the major product when 1.5 equiv of TFA was used. Purification by silica gel column chromatography (cyclohexane/EtOAc 3:7) gave the product in 91% yield (314 mg): yellowish solid; mp 168–170 °C (EtOAc); ESI MS (*m*/*z*) 215 [M + H − C₉H₈NH]⁺; IR (cm⁻¹, film) 3297, 3209, 1618; ¹H NMR (DMSO-*d*₆) δ 1.74 (s, 3H), 2.32 (s, 6H), 3.93 (dd, *J*₁ ≈ *J*₂ = 7 Hz, 2H), 4.60 (t, *J* = 7 Hz, 1H), 6.76–6.94 (m, 4H), 7.19 (d, *J* = 7.5 Hz, 2H), 7.36 (d, *J* = 7.5 Hz, 2H), 7.73 (m, 1H), 10.64 (bs, 2H); ¹³C NMR (DMSO-*d*₆) δ 12.7, 23.0, 34.5, 43.6, 111.2,112.4, 118.8, 119.2, 120.3,128.7, 132.4, 136.0, 170.1. Anal. Calcd for C₂₂H₂₃N₃O (345): C, 76.49; H, 6.71; N, 12.16. Found: C, 76.59; H, 6.77; N, 12.18.

Melatonin. In THF (40 mL) was condensed NH₃ (105 mL) at -78 °C followed by addition of Na (970 mg, 42 mmol). A solution of 3r (2.25 g, 7 mmol) in THF (40 mL) was then added dropwise, and the mixture was stirred at -33 °C for 1 h. Solid NH₄Cl was added, the bath removed, and the ammonia allowed to evaporate. The solvent was evaporated under reduced pressure, water (80 mL) was added, and the aqueous layer was extracted three times with EtOAc (60 mL). The combined organic layers were dried over Na2SO4, filtered, and concentrated. The residue was purified by flash chromatography on silica gel (EtOAc) to give melatonin as a white solid: yield 92% (1.18 g); mp 117–120 °C; ESI MS (m/z): 233 $[M + H]^+$; IR $(cm^{-1}, film)$ 1627; ¹H NMR (CDCl₃) δ 1.93 (s, 3H), 2.94 (t, J = 7 Hz, 2H), 3.59 $(dd, J_1 \approx J_2 = 7 Hz, 2H), 3.86 (s, 3H), 5.69 (bs, 1H), 6.87 (d, J = 8.5)$ Hz, 1H), 7.02 (d, J = 5.0 Hz, 2H), 7.27 (d, J = 8.5 Hz 1H), 8.32 (bs, 1H); ¹³C NMR (CDCl₃) δ 23.4, 25.3, 39.8, 55.9, 100.4, 112.1, 112.4, 112.6, 122.7, 127.7, 131.6, 154.1, 170.2. Anal. Calcd for C₁₃H₁₆N₂O₂ (232): C, 67.22; H, 6.94; N, 12.06. Found: C, 67.15; H, 6.83; N, 12.26. Data was consistent with that reported in the literature.¹⁸⁶

ASSOCIATED CONTENT

Supporting Information

Experimental details of the reactions and copies of ¹H NMR and ¹³C NMR spectra for all substrates and products. This

material is available free of charge via the Internet at http:// pubs.acs.org.

AUTHOR INFORMATION

Corresponding Author

*E-mail: giovanni.piersanti@uniurb.it.

Notes

The authors declare no competing financial interest.

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